

here, the overvoltage for the production of hydrogen at the tungsten electrode is 0.85 V ( $\text{H}_2\text{O}$ , pH 7.8, 0.1 M  $\text{Na}_2\text{SO}_4$ ), and dihydrogen evolution becomes rapid at  $-1.7$  V (vs. SCE). We use a mixture of  $\text{Lip}^{\text{ox}}$  and  $\text{DTT}^{\text{ox}}$  in Scheme I to increase the overall rate of cathodic disulfide reduction.  $\text{DTT}^{\text{ox}}$  is reduced approximately twice as rapidly as  $\text{Lip}^{\text{ox}}$  at  $-1.0$  V, and a major pathway in the reductions of a mixture of the two disulfides is one in which  $\text{DTT}^{\text{ox}}$  is converted to  $\text{DTT}^{\text{red}}$ , and this  $\text{DTT}^{\text{red}}$  in turn reduces  $\text{Lip}^{\text{ox}}$  to  $\text{Lip}^{\text{red}}$  by thiol-disulfide interchange.<sup>8</sup>  $\text{DTT}^{\text{red}}$  is not a substrate for LipDH.

We illustrate the operation of the regeneration sequence summarized in Scheme I with the synthesis of L-lactate from pyruvate. A 2-L, three-necked, round-bottomed flask containing a stirring bar and attached to an argon line was used as the reaction vessel. The working electrode was 40 ft of coiled tungsten wire<sup>9</sup> (0.030-in. diameter, surface area approximately 300  $\text{cm}^2$ ) and the reference electrode was an unexceptional SCE. The counter electrode was 3 ft of coiled platinum wire (0.040-in. diameter), separated from the working solution by a porous ceramic Soxhlet extraction thimble (VWR) inserted in the center neck of the flask. During electrolysis, the solution in the anode compartment was purged with a slow stream of Ar to remove any  $\text{O}_2$  formed.<sup>10</sup> The flask was charged with 500 mL of imidazole (Im)- $\text{H}_2\text{SO}_4$  buffer (50 mM, pH 7.8). D,L-Lipoamide (2.05 g, 10 mmol),<sup>11</sup>  $\text{DTT}^{\text{ox}}$  (1.52 g, 10 mmol), sodium pyruvate (0.55 g, 5 mmol), and xanthine (0.3 g, 2 mmol)<sup>12</sup> were added, and the solution was readjusted to pH 7.8. LipDH (EC 1.6.4.3, from torula yeast) and L-LDH (EC 1.1.1.27) were coimmobilized in PAN gel:<sup>13</sup> 140 mL of swollen gel added to the reactor contained 340 U ( $\mu\text{mol min}^{-1}$ ) of LipDH and 460 U of L-LDH. NAD (0.14 mmol) was added, the potential of the tungsten electrode was adjusted to  $-1.0$  V (vs. SCE), and sodium pyruvate (14.3 g, 130 mmol, in 100 mL of Im- $\text{H}_2\text{SO}_4$  buffer) was added at 1.5  $\text{mmol h}^{-1}$ . Reaction was complete in 3.5 days; under these conditions the cathodic reduction of disulfides was overall rate limiting, at least at the start of the reaction.<sup>14</sup> The gel was allowed to settle, the supernatant decanted, and the lactic acid produced isolated as its zinc salt as described previously<sup>15</sup> (13.8 g, 96% pure, 115 mmol, 85% yield based on pyruvate, 96% ee). The turnover numbers<sup>16</sup> (and recovered activities) of the components were as follows: LipDH,  $2.6 \times 10^7$  (88%); L-LDH,  $3.6 \times 10^7$  (90%); NAD, 920; Lip, 13. No effort was made to recover Lip or NAD. Only approximately 5% of the NAD(H) originally added to the reaction mixture remained active at the

conclusion of the reaction; most of the Lip was still present in active form.

This synthesis demonstrates one practical procedure for the electrochemical regeneration of NAD. The efficient reduction of stable disulfides to strongly reducing dithiols represents a new electrochemical reaction and should be useful in other areas of enzymology, especially in protection of enzymes against autoxidation. The isolation of enantiomerically enriched product establishes that the ultimate reduction step—pyruvate to L-lactate—is enzymatic. The immobilization of the enzyme seems to be important to the success of the procedure: immobilization protects the enzymes against deactivation at the electrode surface and protects the electrode surface against poisoning by adsorbed proteins.

The present limitations of this regeneration procedure are that the system is specific for NAD,<sup>17</sup> that the turnover number achieved for NAD is lower by factors of 2–5 than those observed in comparable preparations of L- or D-lactic acid using purely enzymatic regeneration systems<sup>15,18,19</sup> (probably because of electrochemical reduction of NAD to a biologically-inactive product), and that, in common with many electrochemical syntheses, the reaction is complicated by the equipment required.

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**Registry No.** NADH, 58-68-4; NAD, 53-84-9.

(17) Williams, C. H., Jr., In "The Enzymes", 3rd ed.; Academic: New York, 1976; Vol. 13, pp 106-129.

(18) Wong, C.-H.; Daniels, L.; Orme-Johnson, W.; Whitesides, G. M. *J. Am. Chem. Soc.*, manuscript submitted ( $\text{H}_2$ /hydrogenase).

(19) Shaked, Z.; Whitesides, G. M. *J. Am. Chem. Soc.* 1980, 102, 7104-7105 (formate/formate dehydrogenase).

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(8) Szajewski, R. P.; Whitesides, G. M. *J. Am. Chem. Soc.* 1980, 102, 2011-2026. The equilibrium constant for reduction of NAD to NADH by  $\text{DTT}^{\text{red}}$  is  $\sim 1.3$ .

(9) Molybdenum is also an effective cathode for disulfide reduction but gives lower current efficiencies than tungsten.

(10) The anodic products of reaction were not identified.

(11) The presence of  $\text{Lip}^{\text{ox}}$  in large excess (5–10 times the solubility of  $\text{Lip}^{\text{ox}}$  in the buffer employed) resulted in greater selectivity of the tungsten cathode against NAD and pyruvate reduction.

(12) Inclusion of nitrogen heterocycles (nicotinamide, nicotinic acid, AMP, adenine, xanthine) in the reaction mixture substantially reduced the rate of cathodic reduction of NAD, probably by competition for sites active for the reduction of heterocycles on the electrode surface (see Bresmahan et al.<sup>1</sup>).

(13) Pollak, A.; Blumenfield, H.; Wax, M.; Baughn, R. L.; Whitesides, G. M. *J. Am. Chem. Soc.* 1980, 102, 6324-6336.

(14) This method of conducting the reaction results in a high steady-state value for  $[\text{NAD}]/[\text{NADH}]$ . NAD is intrinsically more stable than NADH under these solution conditions but more rapidly destroyed by cathodic reduction.<sup>15</sup> We believe that electrochemical reduction limits the lifetime of NAD(H) in this system.

(15) Wong, C.-H.; Whitesides, G. M. *J. Am. Chem. Soc.*, 1981, 103, 4890.

(16) Turnover number = mole of lactate isolated per mole of enzyme (cofactor).

### A Stereoselective Approach to Acyclic Systems via Condensations of $\alpha$ -Lithiosulfinyl Carbanions and Aldehydes

**Summary:** The stereochemical outcome of condensations of  $\alpha$ -lithiosulfinyl carbanions with aldehydes is presented, and demonstrates useful methodology for generating 1,2-asymmetry, as well as construction of 1,3-asymmetric relationships in acyclic systems.

**Sir:** Development of synthetic methodology for relative asymmetric induction has been recognized as a challenging problem in the chemistry of complex acyclic molecules. In part, as an outgrowth of interest in the synthesis of ionophore antibiotics, new methods have recently demonstrated relative asymmetric induction in acyclic systems.<sup>1</sup> Although a number of procedures establish 1,2-asymmetry,<sup>2</sup> construction of 1,3-asymmetric relationships presents

(1) For an excellent review on acyclic stereocontrol, see: Bartlett, P. A. *Tetrahedron* 1980, 36, 2.



